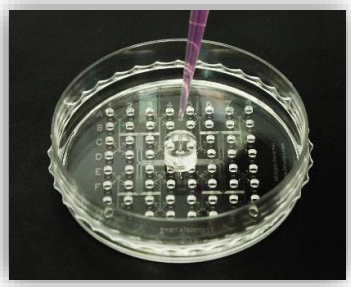
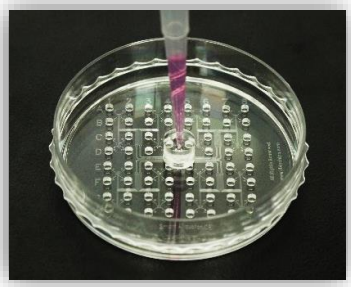
Full Protocol of SACE for Single-Cell Isolation



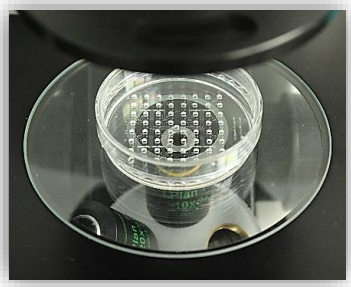
Step 1: Cell Suspension Preparation

1. Harvest adherent or suspension cells and make them into cell suspension.
2. Transfer 2 µl of cell suspension to a Petri Dish.
3. Identify the cell suspension and roughly estimate cell numbers under the microscope.
4. The ideal number of cells is 30-50. If too much/little, dilute/enrich it to match the number.
5. Add 100 µL of PBS/medium to the cell suspension and pipette cells to be fully suspended.
6. Aspirate the 100 µl of cell suspension into pipette tip.



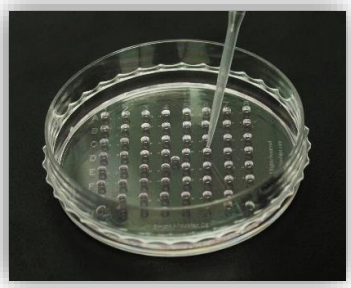
Step 2: Single-Cell Isolation

1. Vertically insert pipette tip into the Inlet Adaptor.
2. Press pipette plunger down (100 µL +P) to aliquot cell suspension into the 64 Outlet Wells.
3. After aliquoting, hold the device and tilt the pipette tip to remove the Inlet Adaptor, without releasing plunger button.



Step 3: Single-Cell Identification

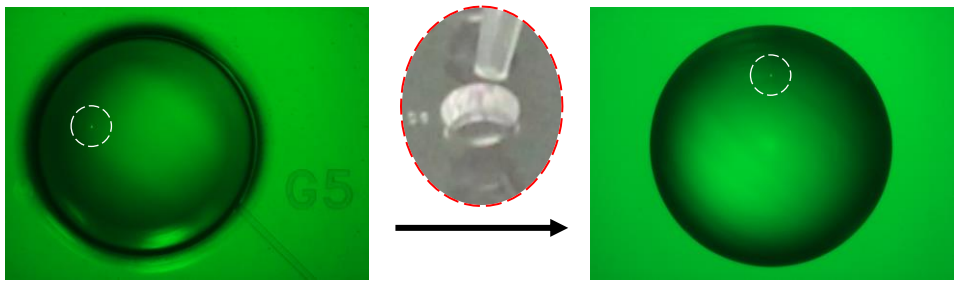
1. Put the SACE under microscope with 10x objective (bright field or fluorescence).
2. Identify and note all Outlet Wells containing only single cells.



Step 4: Single-Cell Transfer

1. Pipette the isolated single cells to be fully suspended within the Outlet Wells.
2. Transfer desired single cells as ~1.5 µL of suspensions to other containers such as PCR tubes.

Before transfer **After transfer**



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